## LABORATORY CALCULATIONS

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## Learning Objectives

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- Understand and be able to use the following
types of calculations:
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- Reference intervals
- Sensitivity/specificity $\qquad$
- ROC curve
- Student $t$ test
- Volume of distribution
- Beer's Law
- Enzyme kinetics $\qquad$
- Basic management calculations
- Buffers $\qquad$
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## Reference intervals

- Validating a reference interval?
- 20 - 60 reference individuals
- Transferring a reference interval?

$$
\text { - } 20 \text { - } \mathbf{6 0} \text { reference individuals }
$$

- Establishing a reference interval
- On a test with well-defined inclusion/exclusion criteria? - a priori sampling - 120 healthy individuals in each partition to get $90 \%$ C.I. at $95^{\text {th }}$ percentile
- On a new analyte? - a posteriori sampling - as many as you can analyze


## Reference intervals

- Establishing a reference interval
-Look at data distribution! - why?
- Example: Chloride on CAVH fluid
- N = 56
- Mean = 101
- Median 100
- SD = 7


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## Reference interval for 3-OH-C16


requency histogram


## 3-OH-C16 reference interval

- $N=197$
- Range = 0.2 - 1.5
- Mean $=0.53 ;$ median $=0.50$
- Non-parametric $95 \%$ reference interval: $\qquad$
- $2.5^{\text {th }}=0.025(198)=4.95=5^{\text {th }}$ value
- $97.5^{\text {th }}=0.975(198)=193^{\text {rd }}$ value
0.3-1.2

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## Transferring a reference interval


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Clinical validity / utility of a test: $\qquad$ sensitivity/specificity/predictive values

- Specificity: the frequency of a negative test when no disease is present; ability to rule out a disease

$$
\text { Spec. }=\frac{T N}{T N+F P} \quad \times 100=\quad(\%)
$$

- Sensitivity: the frequency of a positive test when disease is present; ability of test to detect disease

$$
\text { Sens. }=\frac{T P}{T P+F N} \times 100=\quad(\%)
$$

## Sensitivity/specificity

$$
\text { Spec. }=\frac{\mathrm{TN}}{\mathrm{TN}+\mathrm{FP}} \times 100=(\%) \quad \text { Sens. }=\frac{\mathrm{TP}}{\mathrm{TP}+\mathrm{FN}} \times 100=(\%)
$$

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| Sensitivity/specificity |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Spec. = | TN + FP | $x 100=\quad \text { (\%) }$ | $\text { Sens. }=\mathrm{TP}$ |  | X $100=(\%)$ |
| 3-OHFAs data - good test for diagnosing LCHAD and SCHAD? |  |  |  |  |  |
| SCHAD |  |  | LCHAD |  |  |
|  | SCHAD | $\begin{gathered} \mathrm{No} \\ \text { SCHAD } \end{gathered}$ |  | LCHAD | $\begin{array}{\|c} \mathrm{No} \\ \text { LCHAD } \end{array}$ |
| Positive | 6 (TP) | 15 (FP) | Positive | 8 (TP) | 0 (FP) |
| Negative | 0 (FN) | 182 (TN) | Negative | 0 (FN) | 197 (TN) |
| Spec for SCHAD $=182 / 197 \times 100=92.4 \%$ Spec for LCHAD $=197 / 197 \times 100=100 \%$ |  |  |  |  |  |
| Sens for SCHAD $=6 / 6 \times 100=100 \%$ |  |  | Sens for LCHAD $=8 / 8 \times 100=100 \%$ |  |  |

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Spec for SCHAD $=182 / 197 \times 100=92.4 \% \quad$ Spec for LCHAD $=197 / 197 \times 100=100 \%$
Sens for SCHAD $=6 / 6 \times 100=100 \% \quad$ Sens for LCHAD $=8 / 8 \times 100=100 \%$
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## Clinical/diagnostic utility

- Positive predictive value (PPV) - predictive value of a positive test $\qquad$
PPV $=\quad \frac{\text { TP }}{\text { TP + FP }} \quad \mathbf{X 1 0 0}=\quad \%$

For SCHAD: $6 / 21 \times 100=28.6 \%$ | In general, if prevalence |
| :--- |
| of disease is very low, get |
| more FP, and PPV is bad |

- Negative predictive value (NPV) - predictive value of a negative test


For SCHAD: 199/199 X100 = 100\%
For LCHAD: 197/197 X100 = 100\%
test good for ruling out both disorders
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## ROC curves

- Graphical way to present sensitivity and specificity data
- Software also gives you
- PPV, NPV
- Likelihood ratios: +LR, -LR - likelihood a pos test will be seen in a patient with the disease compared to a patient without the disease
- $\uparrow+L R$ - the better the test is for diagnosing disease
- $\uparrow$-LR - the better the test is at ruling out the disease
- Sensitivity and specificity can be considered reciprocals
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## ROC curves

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AUC $=1.00$
perfect test
$100 \%$ sensitive and specific
AUC $=0.500$
test is no better than
flipping a coin
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To set up a ROC curve

- For each data point, assign a 1 (disorder present) or a 0 , (disorder absent)

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Comparing ROC curves $\qquad$

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## Student test

rcomporng a sande wit me posuason from which was secectiod

$$
t=\frac{\bar{x}-\mu}{s / \sqrt{N}}
$$

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Average age of attendees at a conference is 32
The ages of the 10 attendees in the front row are $35,37,40,30,34,35,38,32$, 34 and 39. Are older attendees more likely to sit on the front row?

$$
\begin{aligned}
t & =(35.4-32) \div(3.13 / \sqrt{ } 10) \\
& =3.4 \div(3.13 / 3.16) \\
& =3.4 / 0.99=3.4243
\end{aligned}
$$

Mean $=35.4$
$s=3.13$
$\mathrm{s}=3.13$
9 degrees of freedom


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## Volume of Distribution $\left(\mathrm{V}_{\mathrm{d}}\right)$

- The Volume of Distribution $\left(\mathrm{V}_{\mathrm{d}}\right)$ is the amount of blood, $\qquad$ per Kg body weight, necessary to contain all of the body burden of drug at equilibrium concentration.

Plasma Concentration $=\frac{\text { Total Body Stores }}{\text { Volume of Distribution }}$
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## Interpreting $\mathrm{V}_{\mathrm{d}}$

- Drugs with low $\mathrm{V}_{\mathrm{d}}$ are contained mostly in the plasma, $\qquad$ because...
- They are highly water soluble (plasma water content is higher than tissues), or
- They are highly protein bound (which prevents them from freely diffusing into tissues
- Drugs with high $\mathrm{V}_{\mathrm{d}}$ are mostly in tissues, and plasma levels may not reflect body burden
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## $\mathrm{V}_{\mathrm{d}}$ calculation

A 175 lb man takes a 5 mg dose of phenobarbital $\left(\mathrm{V}_{\mathrm{d}}=1.0 \mathrm{~L} / \mathrm{Kg}\right)$. What is the maximum plasma phenobarbital concentration you can expect?
Plasma concentration $=$ total body stores $\div$ volume of distribution $\qquad$ $175 \mathrm{lb}=79.4 \mathrm{Kg}$
$C=(5 \mathrm{mg} / 79.4 \mathrm{Kg}) \div 1.0 \mathrm{~L} / \mathrm{Kg}$
$=0.063 \mathrm{mg} / \mathrm{Kg} \div 1.0 \mathrm{~L} / \mathrm{Kg}$
$=0.063 \mathrm{mg} / \mathrm{L}=0.063 \mu \mathrm{~g} / \mathrm{mL}$
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## $\mathrm{V}_{\mathrm{d}}$ calculation

A 55 Kg woman has a plasma theophylline $\left(\mathrm{V}_{\mathrm{d}}=0.5 \mathrm{~L} / \mathrm{Kg}\right)$ concentration of $15 \mu \mathrm{~g} / \mathrm{L}$. What is her total body burden of theophylline?
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Plasma concentration $=$ total body stores $\div$ volume of distribution $\qquad$
$15 \mu \mathrm{~g} / \mathrm{L}=$ (concentration $/ 55 \mathrm{Kg}$ ) $\div 0.5 \mathrm{~L} / \mathrm{Kg}$
$(15 \mu \mathrm{~g} / \mathrm{L})(0.5 \mathrm{Kg} / \mathrm{L})=$ concentration $/ 55 \mathrm{Kg}$
$7.5 \mu \mathrm{~g} / \mathrm{Kg}=$ concentration $/ 55 \mathrm{Kg}$
$\qquad$
$(7.5 \mu \mathrm{~g} / \mathrm{Kg})(55 \mathrm{Kg})=$ concentration
$412.5 \mu \mathrm{~g}$

## Beer's Law

- The mathematical formula that expresses: concentration of an analyte dissolved in solution is directly proportional to it's absorbance.

Caveats:

1) Absorbance must be in the linear range (~0.05-2.0)
2) incident light must be monochromatic
one wavelength
3) no interfering substances may be present absorbances are additive

| Beer's law |  |
| :---: | :---: |
| $A=a b c$ |  |
|  | $\begin{aligned} & \mathrm{A}=\text { absorbance } \\ & \mathrm{a}=\text { absorptivity coefficient } \\ & (\varepsilon=\text { molar units }) \\ & \mathrm{b}=\text { path length of light } \\ & \text { through sample } \\ & \mathrm{c}=\text { concentration } \end{aligned}$ |

## Beer's Law

$\beta-\mathrm{OH}-$ butyrate $+\mathrm{NAD}^{+} \stackrel{3-\mathrm{HBD}}{\longleftrightarrow}$ acetoacetate + NADH + $\mathrm{H}^{+}$ $\qquad$

- 0.1 mL sample added to 2.7 mL buffer, $0.15 \mathrm{~mL} \mathrm{NAD}^{+}$(27 $\mathrm{mmol} / \mathrm{L})$, and $50 \mu \mathrm{~L} 3-\mathrm{HBD}$
( 3 mL total volume: $0.1+2.7+0.15+0.05$ )
- Measured absorbance of produced NADH relative to a blank at 340 nm in a 1 cm cell
$\mathrm{A}=0.57$
- Calculate the $\beta-\mathrm{OH}$-butyrate concentration
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## $\beta$-OH-butyrate

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- $A=a b c$
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$\mathrm{A}=$ absorbance $=0.57$
$\varepsilon=$ molar absorptivity of NADH $\qquad$
$=6.22 \times 10^{3} \mathrm{~L} . \mathrm{mol}^{-1} . \mathrm{cm}^{-1}$
$b=$ path length of light $\qquad$
$c=$ concentration
$0.57=6.22 \times 10^{3} \times 1 \times c$


## $\beta$-OH-butyrate

$0.57=6.22 \times 10^{3} \times 1 \times c$
$c=0.57 \div\left(6.22 \times 10^{3}\right)=9.2 \times 10^{-5} \mathrm{~mol} / \mathrm{L}$

Convert to mmol/L (multiply x $10^{3}$ ) $=0.092 \mathrm{mmol} / \mathrm{L}$
$=\beta$-OH-butyrate in final mixture!
Calculate $\beta$-OHB in sample by multiplying by dilution factor $\left(\mathrm{V}_{\mathrm{T}} / \mathrm{V}_{\mathrm{s}}\right)$

## $\beta$-OH-butyrate

Total volume $=2.7+0.1+0.15+0.05=3.0 \mathrm{~mL}$
$(0.092 \mathrm{mmol} / \mathrm{L} \times 3.0 \mathrm{~mL}) \div 0.1 \mathrm{~mL}$
$=2.76 \mathrm{mmol} / \mathrm{L} \beta-\mathrm{OH}$-butyrate in the sample

Can do this in a single calculation
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## $\beta$-OH-butyrate

Can do this in a single calculation $\qquad$
$c=\frac{(0.57)\left(10^{3}\right)(3.0)}{\left(6.22 \times 10^{3}\right)(0.1)}$

Careful to include all dilution factors and unit conversion factors


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| LD |  |  |  |
| :--- | :--- | :---: | :---: |
| time | Abs |  |  |
| 0 | 0.081 |  |  |
| 1 | 0.114 |  |  |
| 2 | 0.146 |  |  |
| 3 | 0.177 |  |  |
| 4 | 0.211 |  |  |
| 5 | 0.243 |  |  |
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## LD

- $(\Delta \mathrm{Abs} / \varepsilon \times \mathrm{d})\left(10^{6}\right)\left(\mathrm{V}_{\mathrm{T}} / \mathrm{V}_{\mathrm{S}}\right)=\mathrm{U} / \mathrm{L}$
$\mathrm{c}=\frac{(0.032)\left(10^{6}\right)(1.05)}{\left(6.22 \times 10^{3}\right)(0.05)}=108 \mathrm{U} / \mathrm{L}$ $\left(6.22 \times 10^{3}\right)(0.05)$ $\qquad$
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## Enzyme Kinetics

- Kinetics = mathematical description of a reaction as it is happening
- Michaelis and Menten developed a simple model for examining the kinetics of enzyme catalyzed reactions - ASSUMING:


## $E+S \leftrightarrow E S \rightarrow E+P$

formation of ES is reversible
formation of $\mathrm{E}+\mathrm{P}$ is irreversible

- Michaelis-Menten plot
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## Enzyme Kinetics

- Velocity (rate) of the reaction:
- at low [S]:
~straight line;
1st order with respect to [S]
-velocity depends on [S]
- at high [S]:
flat line
zero order with respect to [S]
Rate won't 介 no matter [S]
 unless enzyme concentration $\uparrow$ -velocity depends on enzyme concentration

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| Enzyme Kinetics |  |  |
| :---: | :---: | :---: |
| $\underset{(\mu \mathrm{mmolmin})}{\mathrm{V}}$ | [S] (mmoll) | What is $\mathrm{V}_{\text {max }}$ ? |
| 60 | 200 |  |
| 60 | 20 |  |
| 60 | 2 |  |
| 48 | 0.2 |  |
| 45 | 0.15 |  |
| 12 | 0.013 |  |

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| :---: | :---: | :---: |
| Enzyme Kinetics |  |  |
| $v$ | $[\mathrm{~s}]$ |  |
| (umpolmin) | (mmoll) | What is $\mathbf{v}_{\text {max }}$ ? |
| 60 | 200 |  |
| 60 | 20 | 60 umol/min |
| 60 | 2 |  |
| 48 | 0.2 |  |
| 45 | 0.15 |  |
| 12 | 0.013 |  |
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| Enzyme Kinetics |  |  |
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| $\underset{\substack{(\text { unomimin) } \\ 60}}{V}$ | $\begin{gathered} {[\mathrm{S}]} \\ (\text { (mmoll) } \\ 200 \end{gathered}$ | What is $\mathrm{V}_{\text {max }}$ ? |
| 60 | 20 | $60 \mu \mathrm{~mol} / \mathrm{min}$ |
| 60 | 2 |  |
| 48 | 0.2 | What is $\mathrm{K}_{\mathrm{m}}$ ? |
| 45 | 0.15 | [ $]_{\text {at }} 1 / 2 \mathrm{~V}_{\text {max }}$ |
| 12 | 0.013 | [S] at $30 \mu \mathrm{~mol} / \mathrm{min}$ |

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## Enzyme Kinetics

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## Lineweaver-Burk plot

- $\quad$ Since plot of V vs. [S] is not a straight line, it is $\qquad$ difficult to obtain accurate values of Vmax and Km
- The Lineweaver-Burk plot or double reciprocal plot is a linear transformation of the Michaelis-Menten equation

$$
\frac{1}{v}=\left\{\left[\frac{1}{S}\right]\left[\left(\frac{K m}{V} V_{\text {max }}\right]\right\}+\left(\frac{1}{V_{\text {max }}}\right)\right.
$$

This equation yields a straight line
Where: slope $=\mathrm{Km} / \mathrm{Vmax}$, y intercept $=1 / \mathrm{Vmax}$, $x$ intercept $=-1 / \mathrm{Km}$

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## Lineweaver-Burk

| $[\mathrm{S}]$ | $V(\Delta \mathrm{~A})$ |
| :---: | :--- |
| 0.3 mM | 0.020 |
| 0.6 mM | 0.035 |
| 1.2 mM | 0.048 |
| 4.8 mM | 0.081 |

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| $1 /[S]$ | $1 / V$ |
| :--- | :--- |
| 3.33 | 50 |
| 1.67 | 31.7 |
| 0.83 | 20.8 |
| 0.21 | 12.3 |


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## Lineweaver-Burk experiment

$-1 / \mathrm{Km}=-0.7$
$-1 /-0.7=1.43$
$\mathrm{Km}=1.43 \mathrm{mM}$
$1 / \mathrm{Vmax}=10$
$1 / 10=0.1$

Vmax $=0.1(\mathrm{abs} / \mathrm{min})$


## Management - service contract?

- 75 i-stats at $\$ 9000.00$ each
- Service contract: \$30,000/year/20 I-stats
. $\$ 70,000$ to cover all 75 (instead of $\$ 82,500$ )
- Replacement cost of $\$ 2500.00$ unit
- You have clumsy nurses and average needing to replace 16 units per year
- Do you need the service contract?
- What's the break even number of i-stats?


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## Management - service contract

- 75 i-stats at $\$ 9000.00$ each
- Service contract: $\$ 30,000 /$ year/ 20 I-stats to cover them, $\$ 70,000$ to
$\qquad$ cover all 75.
- Replacement cost of $\$ 2500.00 / u n i$
- You have clumsy nurses and average needing to replace 16 units per year
$16 \times 2500=\$ 40,000.00$ Don't need a service contract.
$70,000 \div 2500=28$ Unless you start breaking more than 28 iStat/year, don't need a service contract

What if you had to buy a new i-Stat whenever you broke one? $\$ 9000$ rather than $\$ 2500$ per broken i-Stat

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## Management - bring that test in-house?

- Considerations -
- Current cost to send test out
- Current test volume
- Current TAT and perceived needs
- Tech time and workflow
- Instrumentation to run assay
- Newly available, FDA-approved assay on current chemistry platform
- LDT assay on esoteric instrument


## Management - bring that test in-house?

- Considerations -
- Current cost to send test out
- Current test volume
- Current TAT and perceived needs
- Tech time and workflow
- Instrumentation to run assay
- Newly available assay on current chemistry platform
- Yes, unless:
- Volume is so low, won't break even on what the test costs

Costs more on chem platform than sending it out

- Volume is so high will impact workflow


## Management - bring that test in-house?

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- Voriconazole -
- Current cost to send test out - $\$ 150.00 /$ test
- Current test volume - 1000/year
- Current TAT and perceived needs -4 days at best; want at least next day if possible
- Tech time and workflow - limited techs on esoteric equipment
- Instrumentation to run assay
- LDT assay on esoteric instrument - MS/MS assay



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Volume drops?

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Volumes increased 45\%


## Equilibrium constant - $\mathrm{K}_{\mathrm{a}}$

-When a weak acid dissociates it forms an equilibrium between the acid form and the $\mathrm{H}^{+}$and base

$$
\mathrm{HA} \longleftrightarrow \mathrm{~A}^{-}+\mathrm{H}^{+}
$$

That equilibrium can be described by a constant $\left(K_{a}\right)$ as: $\qquad$

$$
\mathrm{K}_{\mathrm{a}}=\frac{\left[\mathrm{H}^{+}\right]\left[\mathrm{A}^{-}\right]}{[\mathrm{HA}]}
$$

$\qquad$
Henderson-Hasselbalch equation

$$
-\log \left[\mathrm{H}^{+}\right]=-\log \mathrm{K}_{\mathrm{a}}-\log [\mathrm{HA}]
$$

$$
\overline{[\mathrm{A} \cdot]}
$$

$$
\mathrm{pH}=\mathrm{pK}_{\mathrm{a}}+\log \frac{\left[\mathrm{A}^{-}\right]}{[\mathrm{HA}]} \rightarrow \text { base }
$$

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This is the Henderson-Hasselbalch equation
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$$
\begin{aligned}
\text { If } \mathrm{tCO}_{2}= & 26 \mathrm{mM} \text { and } \mathrm{pCO}_{2}=37.7 \mathrm{~mm} \mathrm{Hg}, \text { what is } \mathrm{pH} \text { ? } \\
\mathrm{pH} & =6.1+\log \frac{26-(0.03)(37.7)}{(0.03)(37.7)} \\
\mathrm{pH} & =6.1+\log \frac{26-1.13}{1.13} \\
\mathrm{pH} & =6.1+\log \frac{24.87}{1.13} \\
\mathrm{pH} & =6.1+\log 22.01 \\
\mathrm{pH} & =6.1+1.34=7.44
\end{aligned}
$$



## Acid/Base Ratio needed to make a buffer

A phosphate buffer, pH 5.7, using dibasic $\qquad$ and monobasic phosphates, $\mathrm{pK}=6.7$ $\left(\mathrm{HPO}_{4}^{-2} / \mathrm{H}_{2} \mathrm{PO}_{4}^{-1}\right)$ (base/acid) $\qquad$

$$
5.7=6.7+\log \frac{\left[\mathrm{HPO}_{4}^{-2}\right]}{\left[\mathrm{H}_{2} \mathrm{PO}_{4}^{-1}\right]}
$$

$5.7-6.7=\log$ of the ratio
$-1=\log$ of ratio (take antilog of both sides of equation)
$0.1=$ ratio $=1: 10$

## Make a buffer

A 150 mM citrate buffer, pH 5.2 ; given: $\mathrm{pK}=4.77$, citric acid MW = 192.12, Na citrate MW = 215.12

$$
\begin{aligned}
& \mathrm{pH}=\mathrm{pK}+\log \frac{\text { [base }]}{[\text { acid }]} \quad 5.2=4.77+\log \frac{[\text { base }]}{[\text { acid }]} \\
& 0.43=\log \frac{\text { base] }}{\text { [acid] }} \\
& 2.69=\text { ratio of base to acid } \\
& \text { so need: } \quad 2.69 \text { moles } / \mathrm{L} \text { base : } 1 \text { mole } / \mathrm{L} \text { acid }
\end{aligned}
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$\qquad$


